

# An overview of Novel and Conventional Methods to Detect Microparticles and Exosomes

Edwin van der Pol



September 23rd, 2010



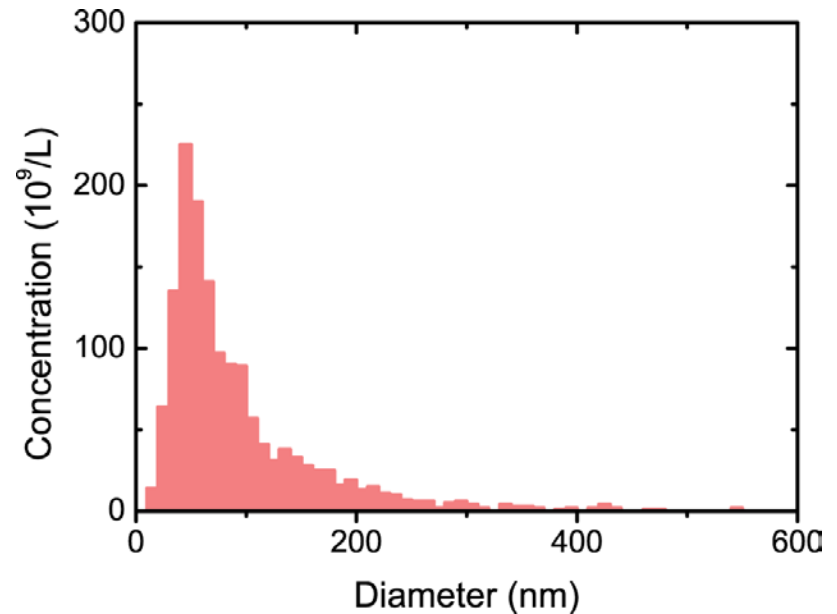
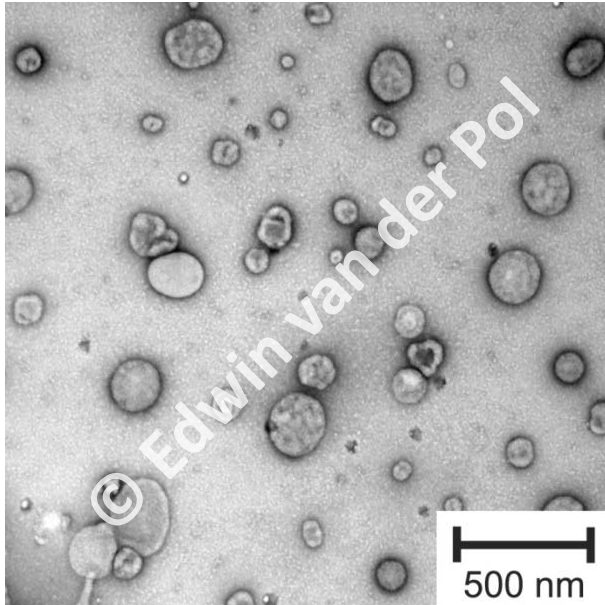
**Academic Medical Center (AMC)**

**University of Amsterdam (UvA)**

Laboratory Experimental Clinical Chemistry (Rienk Nieuwland)

Biomedical Engineering & Physics (Ton van Leeuwen)

# Introduction



- body fluids contain cell-derived vesicles
- clinically relevant information
- problem: vesicle detection



# Outline

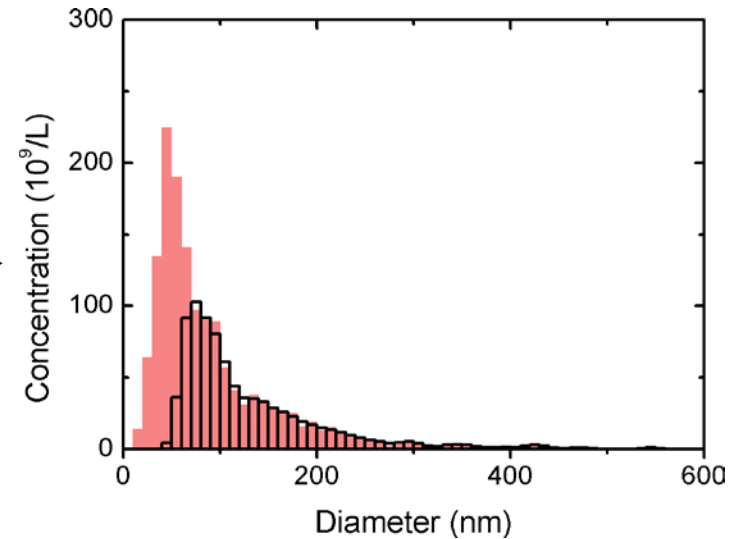
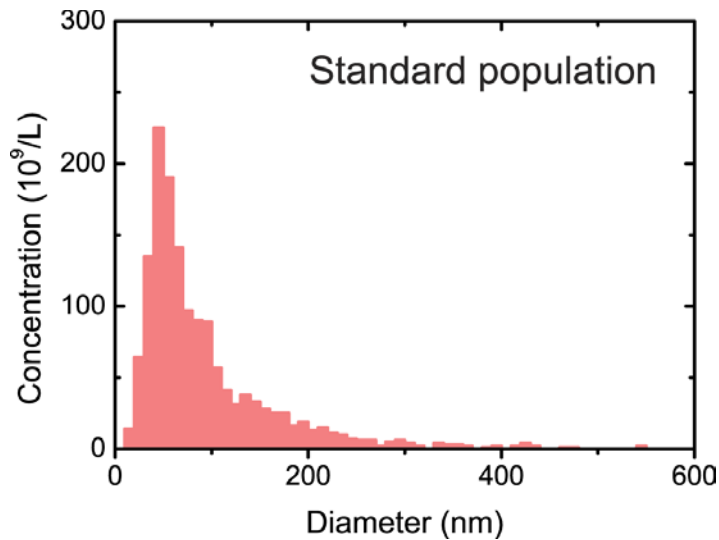
- exploration of detection methods
  - Flow cytometry (FACS)
  - Dynamic Light Scattering (DLS)
  - Nanoparticle Tracking Analysis (NTA)
  - Atomic Force Microscopy (AFM)
  - Impedance-based flow cytometry
- future developments
- conclusions

# Approach: estimate capabilities of methods considering well-known limitations

## Example 1

Detection limit: **50 nm**

Size resolvability: **20 nm**

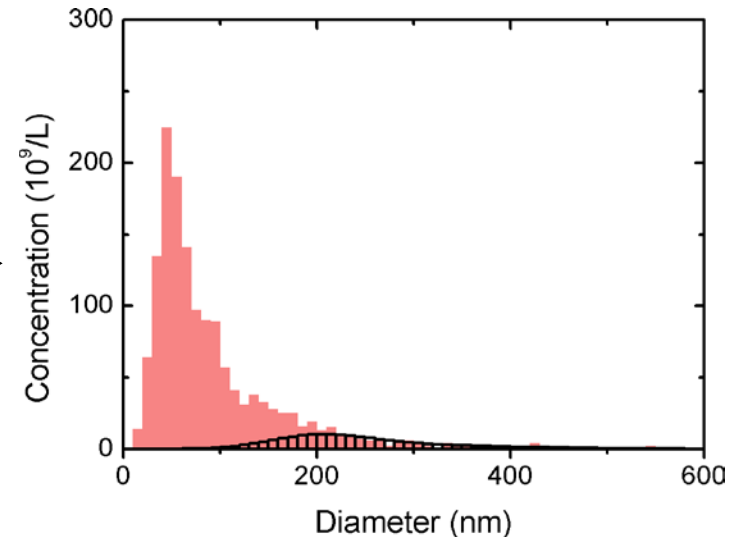
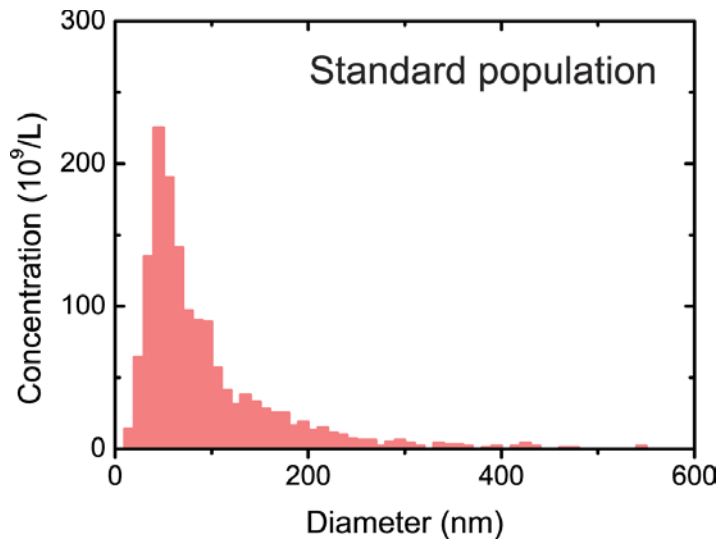


# Approach: estimate capabilities of methods considering well-known limitations

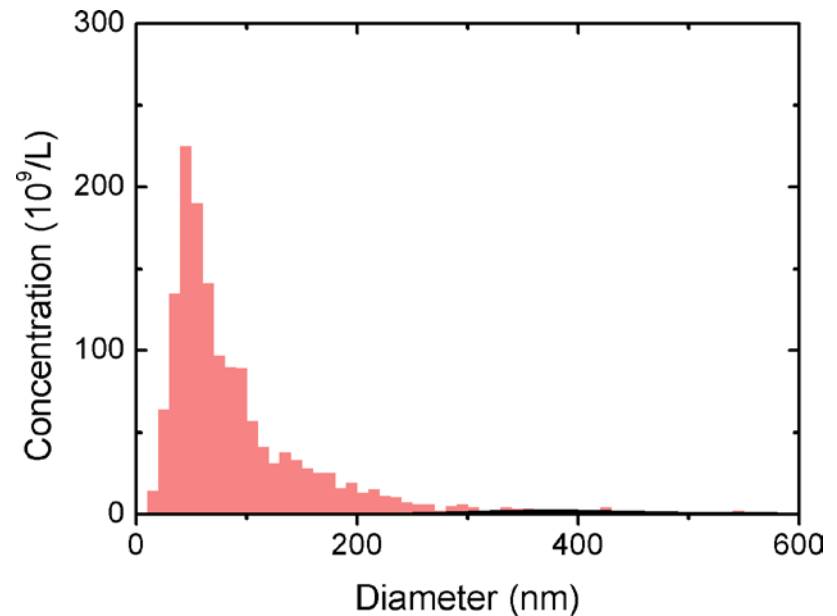
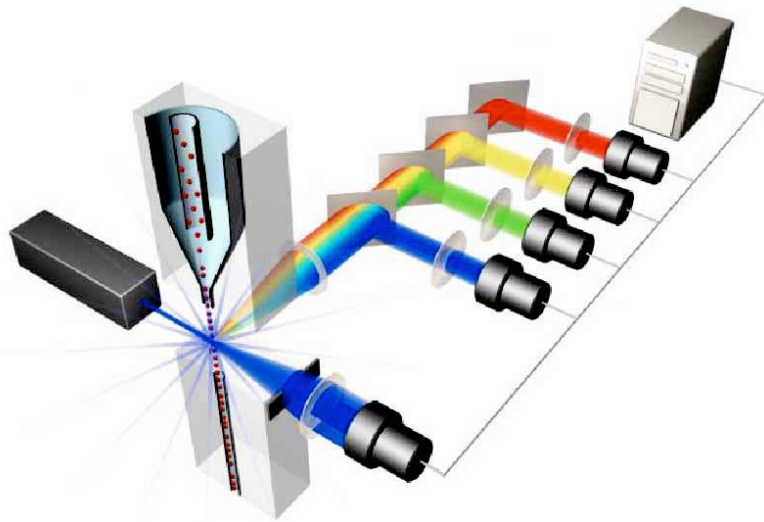
## Example 2

Detection limit: **150 nm**

Size resolvability: **100 nm**



# Flow cytometry (FACS)

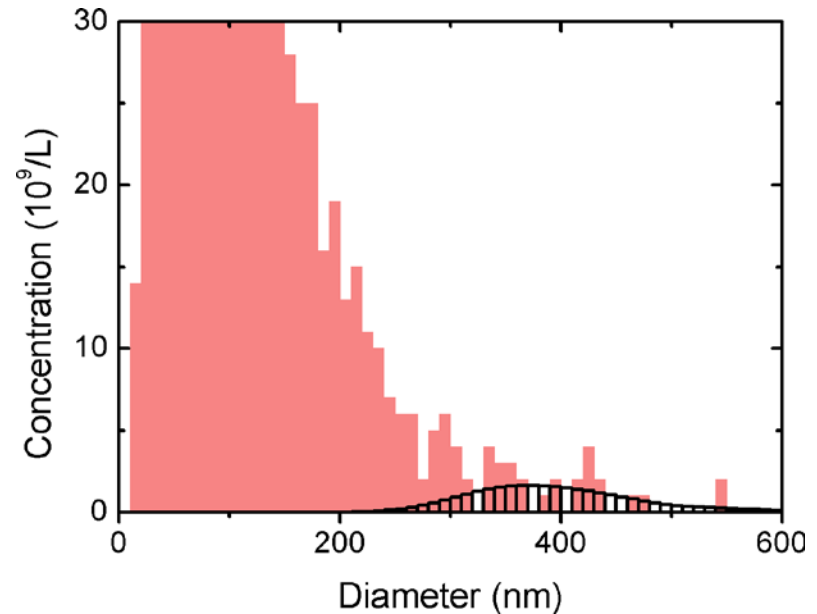
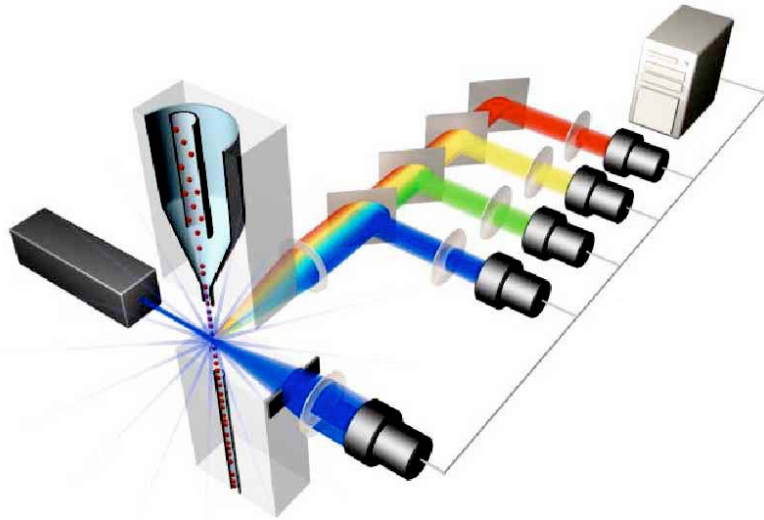


- developed for cell detection ( $>1 \mu\text{m}$ )
- smallest detectable polystyrene bead ( $n=1.6$ ):  $\sim 300 \text{ nm}^{1,2}$
- detection efficiency of vesicles ( $n \approx 1.4$ ) by FACS:  $< 2\%$

1. Robert S. et al. J. Thromb. Haemost. 2009; 7: 190-7

2. Perez-Pujol. et al. S. Cytom. Part. A. 2007; 71: 38-45

# Flow cytometry (FACS)



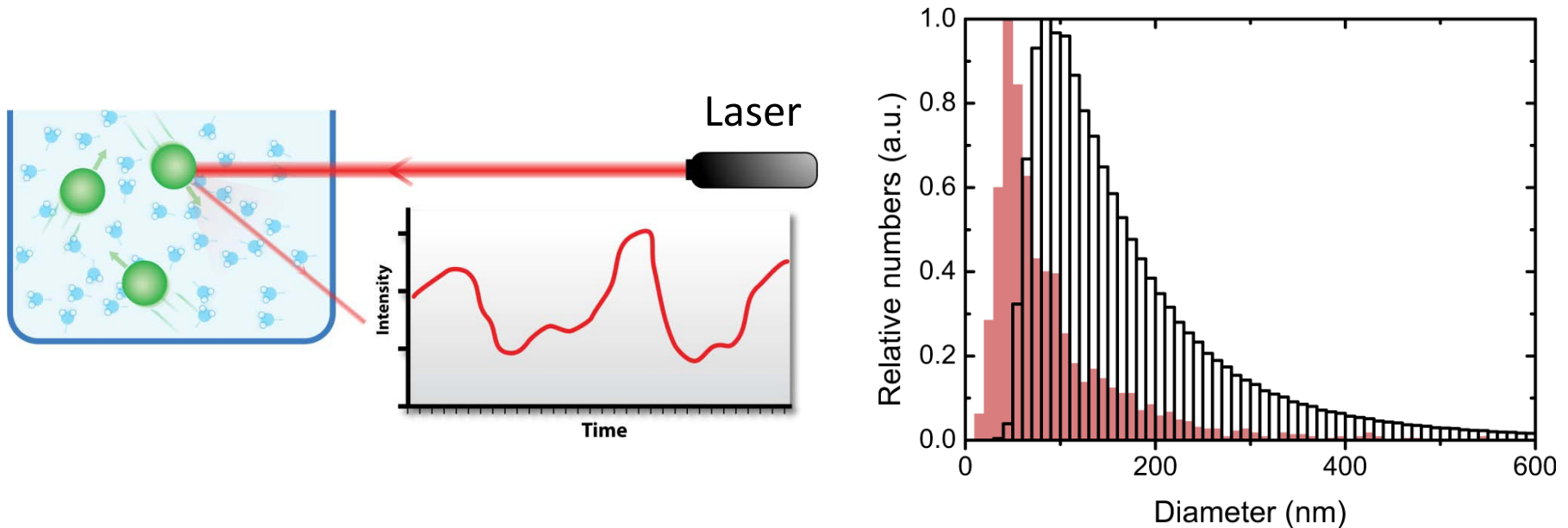
- developed for cell detection ( $>1 \mu\text{m}$ )
- smallest detectable polystyrene bead ( $n=1.6$ ):  $\sim 300 \text{ nm}^{1,2}$
- detection efficiency of vesicles ( $n \approx 1.4$ ) by FACS:  $< 2\%$

1. Robert S. et al. J. Thromb. Haemost. 2009; 7: 190-7

2. Perez-Pujol. et al. S. Cytom. Part. A. 2007; 71: 38-45

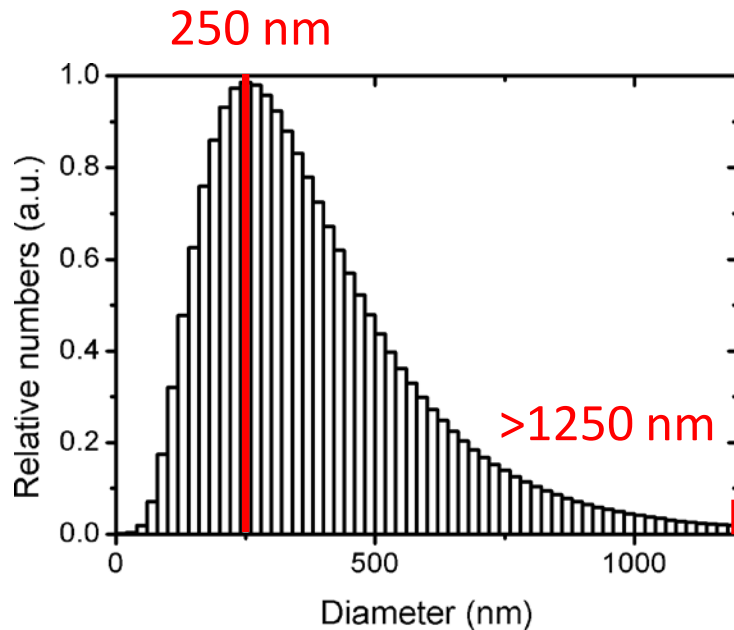


# Dynamic Light Scattering (DLS)

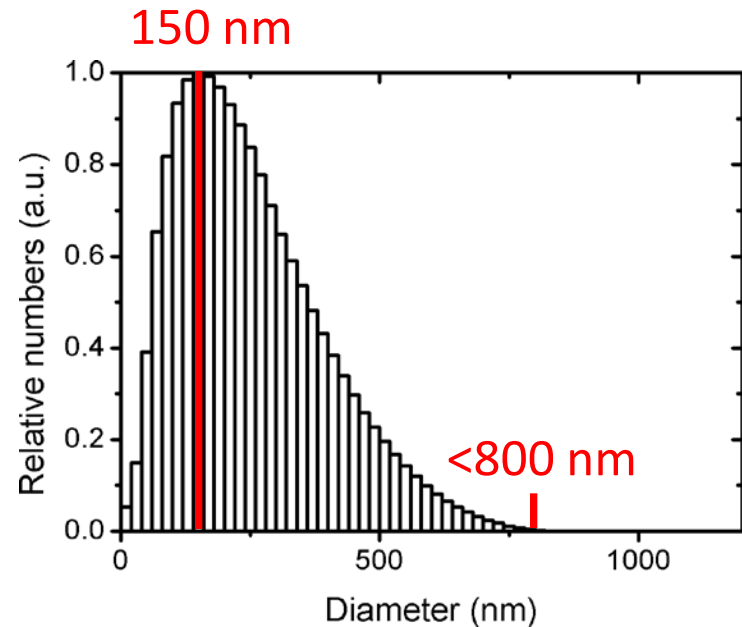


- Brownian motion depends on vesicle diameter
- determines mean *size* of vesicles in fluids
- difficulty with polydisperse samples
- result strongly depends on mathematical algorithm

# DLS applied to vesicles



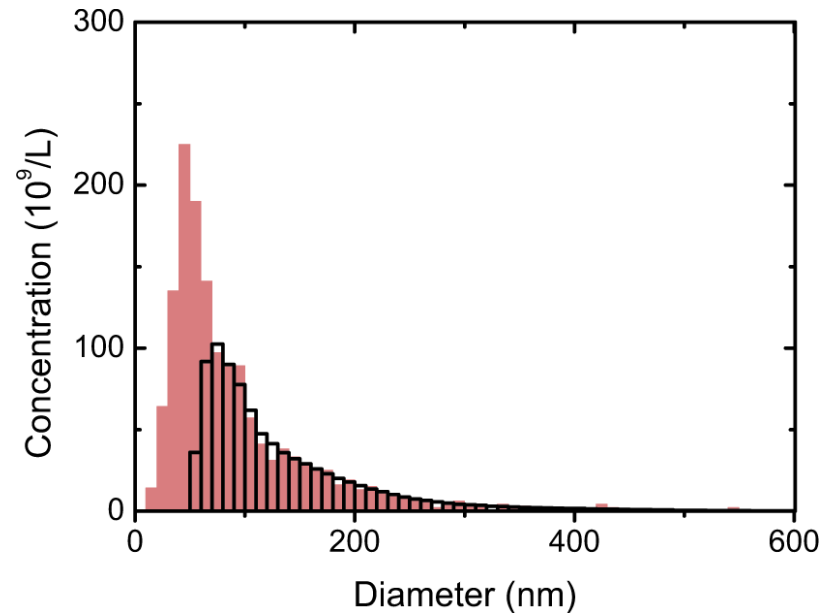
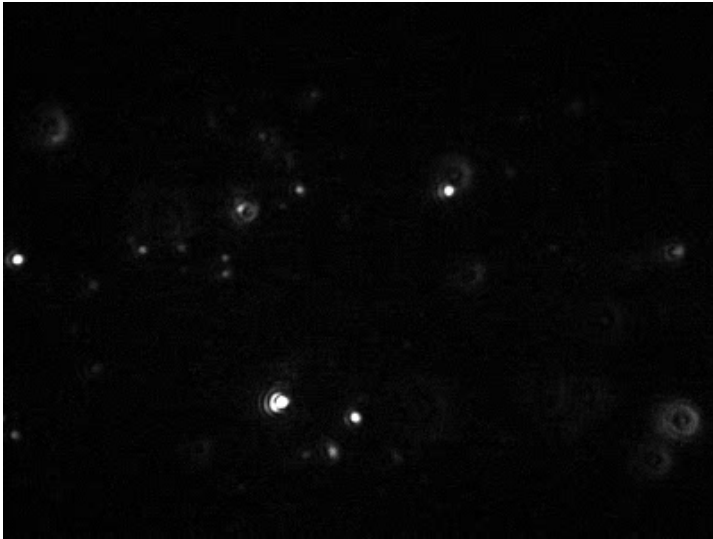
N5 Submicron Particle Size  
Analyser (Beckman Coulter)<sup>3</sup>



Zetasizer Nano S  
(Malvern Instruments Ltd)<sup>3</sup>

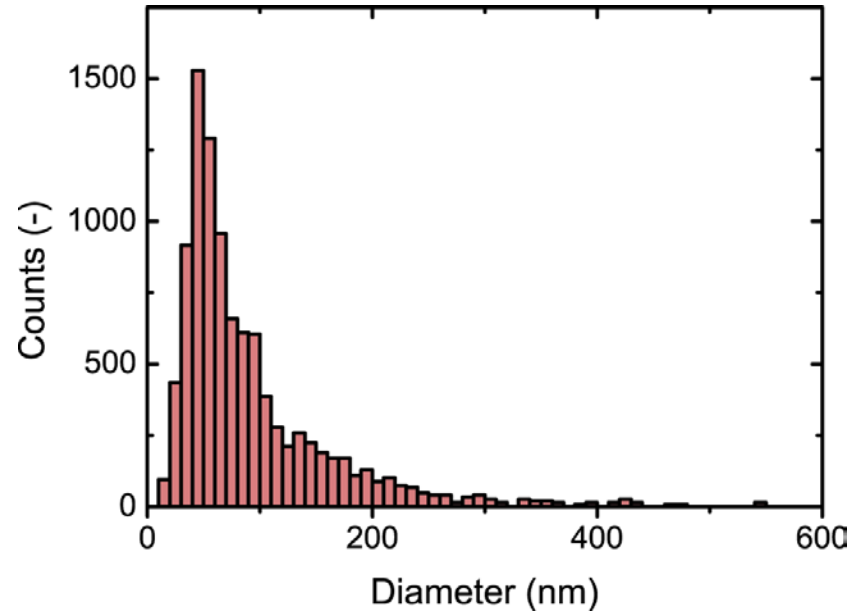
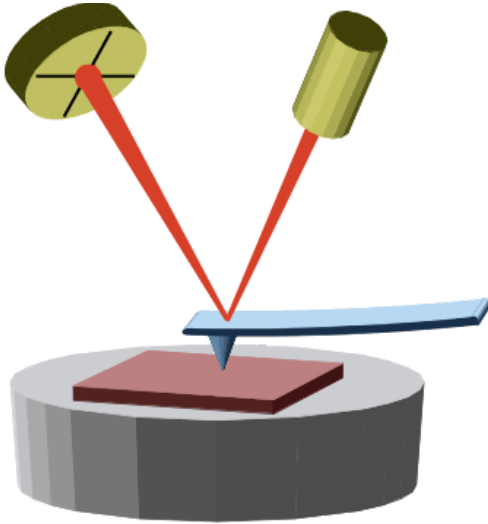
- results are system dependent
- no determination of absolute *size* and *concentration*

# Nanoparticle Tracking Analysis (NTA)



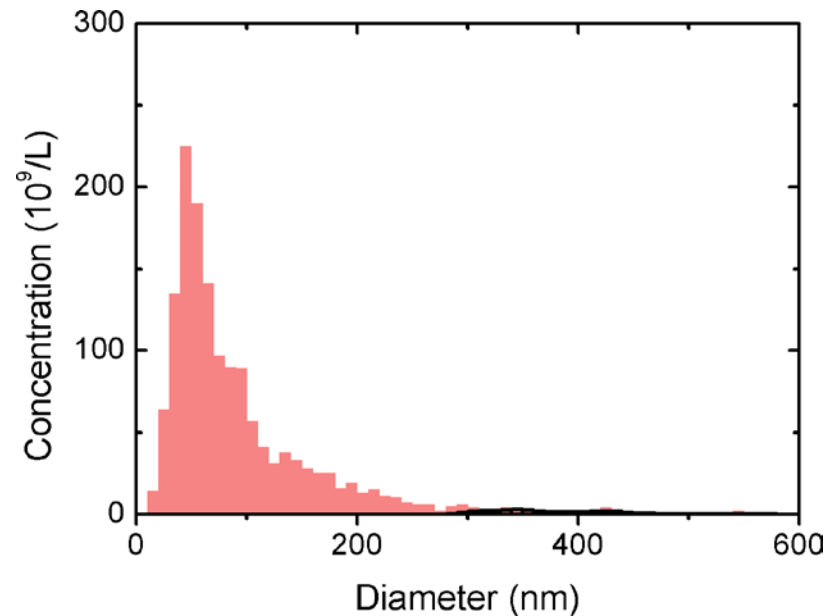
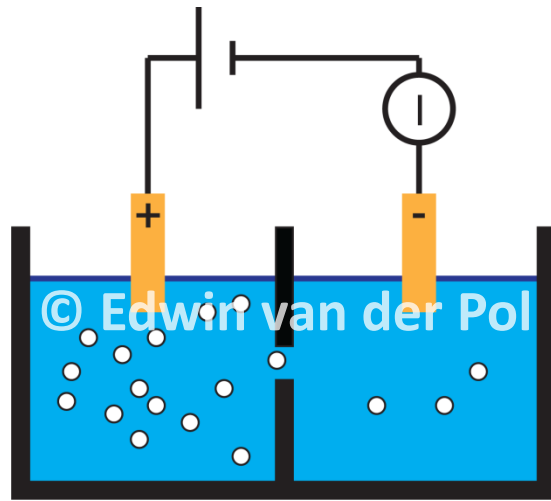
- determines *size* and *concentration* of vesicles in fluids<sup>4</sup>
- present detection limit: ~50 nm for vesicles
- can potentially be extended with fluorescence detection

# Non optical methods: Atomic Force Microscopy (AFM)



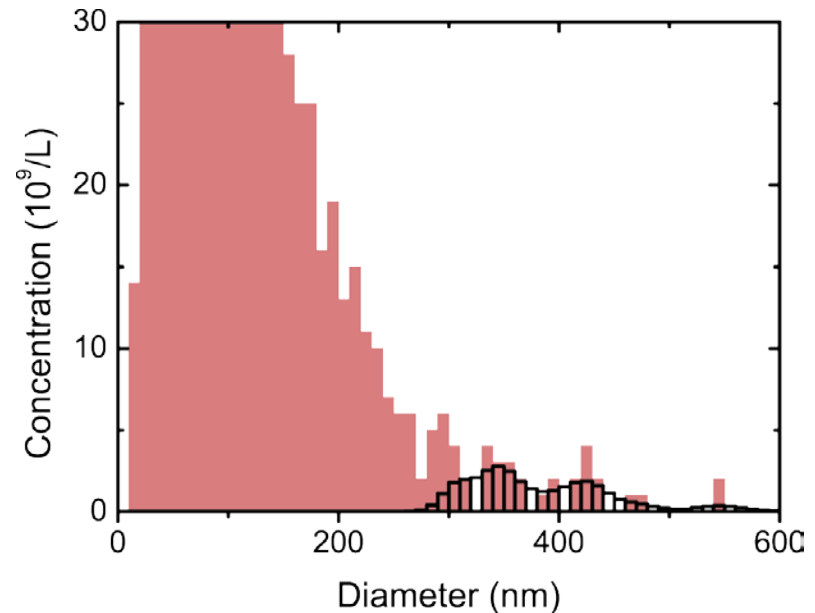
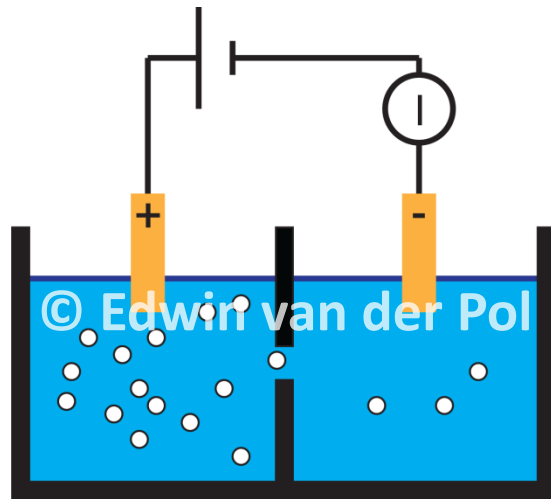
- provides information on *size, concentration, biochemical composition, and cellular origin*<sup>5</sup>
- binding efficiency and influence of binding on vesicle deformation unknown

# Impedance-based flow cytometry



- determines *size* and *concentration* of vesicles
- present detection limit: ~300 nm<sup>6</sup>
- can be combined with flow cytometry

# Impedance-based flow cytometry



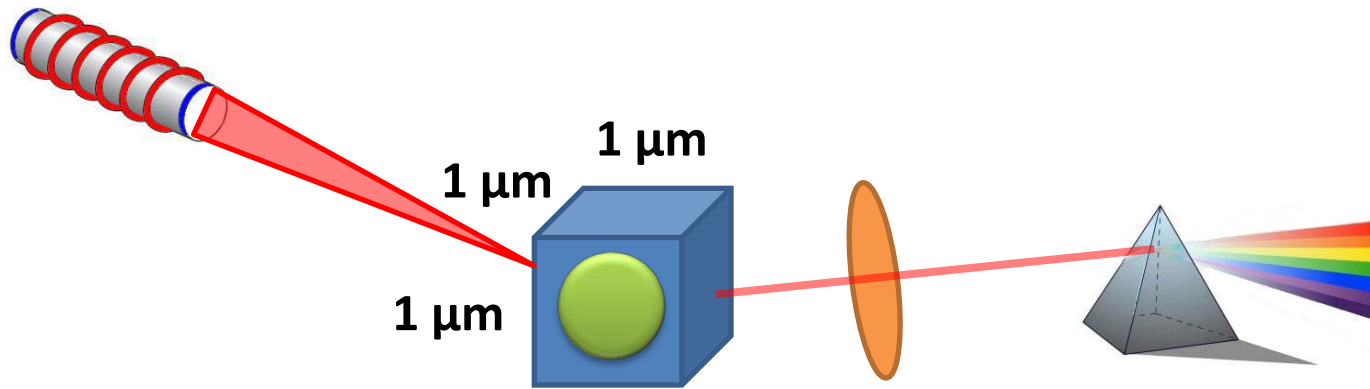
- determines *size* and *concentration* of vesicles
- present detection limit: ~300 nm<sup>6</sup>
- can be combined with flow cytometry

# Overview

Method	Size	Concentration	Biochemical information	Measurement time
Transmission Electron Microscopy (TEM)	✓	✗	☹️	hours
Flow cytometry (FACS)	✗	☹️	✓	seconds
Dynamic Light Scattering (DLS)	☹️	✗	✗	minutes
Nanoparticle Tracking Analysis (NTA)	☹️	☹️	to be investigated	minutes
Atomic Force Microscopy (AFM)	✓	☹️	✓	hours
Impedance-based flow cytometry	✗	☹️	✗	seconds

# Future developments

- Raman microspectroscopy<sup>7</sup>:
  - determine size, concentration, and chemical composition of vesicles in fluids *label-free*





# Conclusions

- vesicle detection remains challenging
- applications of novel and conventional methods requires further investigation