

Supplemental information to:

Centrifugation affects the purity of liquid biopsy-based tumor biomarkers

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Sample properties

Platelets have a log-normal volume distribution (1), and a discoid shape with an aspect ratio of 4-10 (2). For the model, we assumed platelets to be an oblate spheroid with aspect ratio 7, leading to an S for platelets of 2.0 assuming the platelet moves at sufficiently low speeds such that all orientations are equally feasible (3). If this assumption is false, and the platelet is oriented in the energetically most favorable orientation, S would need to be 1.5 (3). Platelets can have a volumetric mass density of 1.05-1.09 g/mL, where the lower density is associated with platelets that have secreted their α -granule content but with a unaffected platelet volume distribution (4, 5). Typical mass densities of EVs determined in Optiprep are found between 1.08 and 1.11. This seems very high compared to typical volumetric mass densities for cells of 1.05-1.08 (6). However, because of the highly skewed particle size distribution this likely pertains to EVs smaller than 200 nm, which have a relatively large membrane content. Modelling an EV as a small cell, i.e. a 8 nm thick membrane consisting of 50% protein (volumetric mass density 1.30), 50% phospholipids (volumetric mass density 1.01) and filled with cytoplasm (volumetric mass density 1.06) (6), would mean that 100 and 200 nm EV have volumetric mass densities of 1.099, and 1.081 respectively. These densities are in good agreement with literature values, while a 1000 nm EV would have a density of 1.064, which is within the range of densities of a cell. Lastly, the mass density of DNA is only available in cesium chloride media, and the length of ccfDNA is unknown. Assuming a length shorter than 1000 basepairs, the ccfDNA would be smaller than 50 nm.

Table S1. Literature values for components in model.

	Property at 20°C, 1 bar	Values in article	Population/sample type	References
CTCs				
Density (g/mL)	1.05 +/- 0.002		lung carcinoma cell line	(7)
	1.056 +/- 0.004		cervical carcinoma cell line	(8)
Diameter (µm)	median 10.7	(measured in range 8-20)	276 castration resistant PCa	(9)
	median 11.0	(measured in range 8-20)	464 colorectal cancer patients	(9)
	median 13.1	(measured in range 8-20)	177 breast cancer patients	(9)
Platelets				
Density (g/mL)	1.05-1.09 @ 4°C	freq mean 1071 SD 6	17 normal donors	(4)
	1.04-1.08 @ 23°C		30 normal donors	(10)
	1.05-1.08 @ RT	freq mean 1067 SD 4.5	15 normal donors	(5)
Diameter (µm)	From spherical approximation of impedance based volume determinations			
	2.5 +/- 0.6 (mean +/- SD)	range 1-21, mean 8.2, SD 6.1 fL	55 normal donors	(11)
	2.3 +/- 0.6	lognormal mean 6.6 SD 4.3 fL	50 normal donors	(1)
Shape factor	“Platelets are discoids 0.5 um thick and 2-5 um diameter” -> aspect ratio 4-10			(2)
	1.55, 2.03, 2.48	aspect ratio 4, 7, 10		(3)
EVs				
Density (g/mL)	Excluded last round culture with exosome containing media, included only iso-osmotic gradient media			
	1.08-1.10		color. carcinoma cell line derived EVs	(12)
	1.09-1.11	sample in 0.25M sucrose before DGC	color. carcinoma cell line derived EVs	(13)
	1.094-1.143		breast cancer cell line derived EVs	(14)
	1.093	some sucrose from previous steps	colorectal carcinoma ascites	(15)
	1.087	some sucrose from previous steps	melanoma in mouse model	(16)

Table S1 (continued)

	Property @ 20°C, 1 bar	Values in paper	Population/sample type	References
ccfDNA				
Density (g/mL)	1.7		in CsCl gradient	(17)
Size (nm)	2 by 304 (1k base pairs)			(6)
PBS				
Density (g/mL)	1.004			(18)
Viscosity (mPa . s)	1.193			(18)
Plasma				
Density (g/mL)	1.026 (CI 1.024-1.027)	@ 4, 25,37 °C	25 normal donors	(19)
	1.0245 (CI 1.0215-1.0262)		32 normal donors	(20)
	1.0253 (CI 1.0217 - 1.0273)		122 pathological donors	(20)
Viscosity (mPa . s)	Converted to 20°C through 2.2% increase per °C (Arrhenius)			(21)
	1.6 - 1.9	1.1 - 1.3 @ 37°C	'normal range'	(22)
	1.70	1.52 @ 25°C	57 normal, no influence of sex, age	(23)
	1.80 (1.75/1.91)	1.24 (25p-75p 1.20-1.31) @ 37°C	Unspecified	(24)

25p-75p: 25-75 percentile, ccfDNA: circulating cell-free DNA, CI: 95% confidence interval, Color.: Colorectal, CTCs: circulating tumor cells, DGC: density gradient centrifugation, EVs: extracellular vesicles, PBS: phosphate-buffered saline, PCa: prostate cancer, RT: room temperature, SD: standard deviation.

References

1. Paulus J-M. Platelet Size in Man. *Blood*. 1975;46(3):321-36.
2. White JG. Platelet Structure. In: Michelson AD, editor. *Platelets*. 2nd ed. London, UK: Elsevier; 2007. p. 45-73.
3. Leith D. Drag on Nonspherical Objects. *Aerosol science and technology*. 1987;6(2):153-61.
4. van Oost BA, Timmermans A, Sixma JJ. Evidence That Platelet Density Depends on the Alpha-Granule Content in Platelets. *Blood*. 1984;63(2):482-5.
5. Corash L, Tan H, Galnick HR. Heterogeneity of Human Whole Blood Platelet Subpopulations. I. Relationship between Buoyant Density, Cell Volume, and Ultrastructure. *Blood*. 1977;49(1):71-87.
6. Milo R, Jorgensen P, Moran U, Weber G, Springer M. Bionumbers—the Database of Key Numbers in Molecular and Cell Biology. *Nucleic acids research*. 2009;38(suppl_1):D750-D3.
7. Bryan AK, Hecht VC, Shen W, Payer K, Grover WH, Manalis SR. Measuring Single Cell Mass, Volume, and Density with Dual Suspended Microchannel Resonators. *Lab on a chip*. 2014;14(3):569-76.
8. Wolff DA, Pertoft H. Separation of Hela Cells by Colloidal Silica Density Gradient Centrifugation: I. Separation and Partial Synchrony of Mitotic Cells. *The Journal of cell biology*. 1972;55(3):579-85.
9. Coumans FA, van Dalum G, Beck M, Terstappen LW. Filter Characteristics Influencing Circulating Tumor Cell Enrichment from Whole Blood. *Plos One*. 2013;8(4):e61770.
10. Chamberlain K, Penington D. Monoamine Oxidase and Other Mitochondrial Enzymes in Density Subpopulations of Human Platelets. *Thrombosis and Haemostasis*. 1988;59(01):029-33.
11. Roper PR, Johnston D, Austin J, Agarwal SS, Drewinko B. Profiles of Platelet Volume Distributions in Normal Individuals and in Patients with Acute Leukemia. *American Journal of Clinical Pathology*. 1977;68(4):449-57.
12. Choi D-S, Choi D-Y, Hong B, Jang S, Kim D-K, Lee J, et al. Quantitative Proteomics of Extracellular Vesicles Derived from Human Primary and Metastatic Colorectal Cancer Cells. *Journal of Extracellular Vesicles*. 2012;1(1):18704.
13. Ji H, Greening DW, Barnes TW, Lim JW, Tauro BJ, Rai A, et al. Proteome Profiling of Exosomes Derived from Human Primary and Metastatic Colorectal Cancer Cells Reveal Differential Expression of Key Metastatic Factors and Signal Transduction Components. *Proteomics*. 2013;13(10-11):1672-86.
14. Baietti MF, Zhang Z, Mortier E, Melchior A, Degeest G, Geeraerts A, et al. Syndecan–Syntenin–Alix Regulates the Biogenesis of Exosomes. *Nature cell biology*. 2012;14(7):677.
15. Choi DS, Park JO, Jang SC, Yoon YJ, Jung JW, Choi DY, et al. Proteomic Analysis of Microvesicles Derived from Human Colorectal Cancer Ascites. *Proteomics*. 2011;11(13):2745-51.
16. Lee E-Y, Park K-S, Yoon YJ, Lee J, Moon H-G, Jang SC, et al. Therapeutic Effects of Autologous Tumor-Derived Nanovesicles on Melanoma Growth and Metastasis. *Plos One*. 2012;7(3):e33330.
17. MATTOCCIA E, COMINGS DE. Buoyant Density and Satellite Composition of DNA of Mouse Heterochromatin. *Nature New Biology*. 1971;229(6):175.

18. Lide D. Dr 1995–1996 Crc Handbook of Chemistry and Physics. Boca Raton, FL: CRC Press.
19. Trudnowski RJ, Rico RC. Specific Gravity of Blood and Plasma at 4 and 37 C. Clinical Chemistry. 1974;20(5):615-6.
20. Kenner T. The Measurement of Blood Density and Its Meaning. Basic research in cardiology. 1989;84(2):111-24.
21. Eckmann DM, Bowers S, Stecker M, Cheung AT. Hematocrit, Volume Expander, Temperature, and Shear Rate Effects on Blood Viscosity. Anesthesia & Analgesia. 2000;91(3):539-45.
22. Késmárky G, Kenyeres P, Rábai M, Tóth K. Plasma Viscosity: A Forgotten Variable. Clinical hemorheology and microcirculation. 2008;39(1–4):243-6.
23. Lawrence J. The Plasma Viscosity. Journal of Clinical Pathology. 1950;3(4):332.
24. Hess EL, Cobure A. The Intrinsic Viscosity of Mixed Protein Systems, Including Studies of Plasma and Serum. The Journal of general physiology. 1950;33(5):511-23.